Correlation of Leukocyte Subtypes, Neutrophil-to-Lymphocyte Ratio, and Functional Outcome in Brain Metastasis

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Abstract

BACKGROUND: As the most cause of death in patients with solid extracranial malignancy, brain metastasis (BM) nowadays being studied extensively especially on how to find a reliable laboratory marker that can correlate with its clinical outcome. Leukocyte subtypes, primarily neutrophils and lymphocytes and its ratio known as Neutrophils-Lymphocyte Ratio (NLR) have been known before its relationship with progression of BM from other solid tumours.

AIM: The objectives of this research to study the correlation of leukocyte subtypes, neutrophil-lymphocyte ratio & functional outcome in brain metastasis.

METHODS: The study subjects were recruited consecutively from the study population. Venous blood was taken 5 ml of venous blood samples done in the first day of admission on emergency department and neurology clinic of Neurology Department of Adam Malik General Hospital before any drug injections. Samples were kept in vacutainer tubes containing ethylenediaminetetraacetic acid (EDTA) and sent to Department of Clinical Pathology laboratory of Adam Malik General Hospital, immediately centrifuged at 3100 rpm for 10 minutes in -20°C temperature and analyzed using Sysmex XT-2000i. Functional outcome of the patient assessed using Karnofsky performance scale (KPS) in a cross-sectional manner with laboratory analysis.

RESULTS: We conduct a mean differences and correlational leukocytes and its subsets analysis of 72 BM patients resulting in significant positive correlation on lymphocyte percentage (r = 0.383, p = 0.001) and lymphocyte absolute (r = 0.265, p = 0.024), also significant negative correlation on neutrophils (r = -0.240, p = 0.042) and NLR (r = -0.432, p < 0.001) with Karnofsky performance scale (KPS).

CONCLUSION: Increased lymphocyte absolute and lymphocyte percentage correlated significantly (p < 0.05) with better KPS, while elevated neutrophils percentage and increased NLR show significant correlation with worse outcome of BM patients.

Introduction

Brain metastasis (BM) is one of the central nervous system consequences of primary extracranial malignancy that mainly happen on 20-40% in adult cancer patients. As one of the main cause of death for these population, its progression is determined by several condition such tumour cell migration (intravasation, dissemination and extravasation) and colonisation also microenvironment inflammation that correlates in the severity of the disease. The basic mechanism of BM begin with Paget’s “seed and soil” hypothesis included three main principles: the heterogeneous population of a tumour with different characteristics, specific traits of metastasis and secondary tumoral microenvironment [1], [2], [3], [4].

The human immune system, in general, have several myeloid cells especially leukocyte subtypes which are responsible in cancer metastasis regulation, such neutrophils and monocytes; both considered immunocytes that could secrete vascular endothelial growth factor (VEGF) contributing to increased tumour size and development. While regulation of aspartic proteinase cathepsin E, which act as tumour suppressor activated by lymphocytes, macrophage and a dendritic cell which and apoptosis induction [5], [6].

Neutrophil-to-lymphocyte ratio (NLR) is the ratio of the absolute neutrophil count to the absolute lymphocyte count and used as one of a simple, rapid, cost-effective and indicative marker of inflammatory process and stress of human body as it can be obtained on complete blood count (CBC) analysis.
While normal immune system would prevent the dissemination of cancer cells and prevent proliferation, about 0.01% escaping circulating tumor cells (CTC) survival from primary sites resulting from shear stress, mechanical detachment even cell-mediated cytotoxicity which provoked epithelial to mesenchymal transition (EMT) regulated by tumor necrosis factor α (TNF-α) and nuclear factor κB (NF-κB). Individual immunocompetence capability could also be estimated by NLR and correlated with disease progression and functional outcome in malignancy measured by Karnofsky performance scale (KPS) [5]. On this basis, elevated NLR ratio could be consequences of either neutrophilia which promotes tumour granulocyte colony-stimulating factor (GM-CSF) and/or lymphopenia that can cause circulating tumour cell escape systemic immunosurveillance in the human body [5], [6].

Growing interest of NLR count on has been established today as several studies tried to elaborate between this ratio and progression of the disease that implicated by even slight process of inflammation, for examples malignancy in various primary organ (e.g.: small cell lung cancer, renal cell carcinoma, pancreatic cancer, primary liver ca, breast cancer, glioblastoma multiforme), arterial disease (e.g: acute coronary syndrome, cerebral infarction, Takayasu arteritis), degenerative disease (e.g: ankylosing spondylitis) also in some autoimmune disorders (e.g: psoriasis, systemic lupus erythematosus [SLE]) [7], [8], [9], [10], [11], [12], [13], [14], [15], [16], [17], [18], [19].

Methods

Ethical approval received from the Health Research Ethical Committee, Faculty of Medicine Universitas Sumatera Utara, Medan-Indonesia. The research was conducted from January 2016 to December 2016 at Adam Malik General Hospital recruiting 72 consecutive sample of patients diagnosed with BM. Confirmation of BM diagnosis was done by means if there are any clinical neurologic deficits and/or any suspicious prove of in any established modalities (laboratory tumour markers, chest radiography, ultrasonography, computed tomography (CT) scan, radionuclide tracing and/or another diagnostic test) supporting central nervous system (CNS) involvement from other extracranial primary malignancy [20].

Exclusion criteria in this study including sepsis, septicaemia, bacteremia, septic shock and other systemic inflammatory response syndrome (SIRS)-like condition, leukemia and leukemoid reaction, prior dexamethasone, methylprednisolone and/or any intravenous steroid injection, autoimmune disease (e.g: SLE, scleroderma) or immunodeficient syndrome (e.g: HIV/AIDS) essentials heart, liver and/or kidney disorders, pneumonia, seizure at the beginning or during hospitalization, massive upper and/or lower GI bleeding which all of the above could modify, primarily suppress the baseline immunological condition of the patients [21], [22], [23].

Informed consent was asked from the patients (or their legal responders in any means the patients cannot give consent) and collection of 5 ml of venous blood samples done in the first day of admission on emergency department and/or neurology clinic of Neurology Department of Adam Malik General Hospital before any drug injections. Samples were kept in vacutainer tubes containing ethylenediaminetetraacetic acid (EDTA) and sent to Department of Clinical Pathology laboratory of Adam Malik General Hospital, immediately centrifuged at 3100 rpm for 10 minutes in -200 C temperature and analysed using Sysmex XT-2000i. Functional outcome of the patient assessed using Karnofsky performance scale (KPS) in a cross-sectional manner with laboratory analysis.

Data were collected and calculated using IBM SPSS Statistic for Windows, Version 24.0 and test of normality was conducted using Kolmogorov-Smirnov on more than 50 study sample. Variables characteristics were shown in Table 1 while mean differences were compared between groups using one-way analysis of variant (ANOVA) (Table 2), and each variable was correlated with KPS using Pearson’s correlation test on the nature of normal distribution of the samples (Table 3).

Table 1: Characteristics of BM Subjects

<table>
<thead>
<tr>
<th>Variable Characteristics (n = 72)</th>
<th>Mean (Percentage)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>51.74 ± 11.17 years old</td>
</tr>
<tr>
<td>Minimum</td>
<td>22 years old</td>
</tr>
<tr>
<td>Maximum</td>
<td>74 years old</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>50 (41.7%)</td>
</tr>
<tr>
<td>Female</td>
<td>42 (58.3%)</td>
</tr>
<tr>
<td>Origins of Primary Tumor</td>
<td></td>
</tr>
<tr>
<td>Pulmonary</td>
<td>37 (51.4%)</td>
</tr>
<tr>
<td>Breast</td>
<td>17 (23.6%)</td>
</tr>
<tr>
<td>Cervix-genito-urinary</td>
<td>10 (13.9%)</td>
</tr>
<tr>
<td>Others</td>
<td>4 (5.5%)</td>
</tr>
<tr>
<td>Unknown</td>
<td>4 (5.5%)</td>
</tr>
<tr>
<td>KPS</td>
<td></td>
</tr>
<tr>
<td>80-100</td>
<td>10 (13.89%)</td>
</tr>
<tr>
<td>50-70</td>
<td>34 (47.22%)</td>
</tr>
<tr>
<td>0-40</td>
<td>28 (38.89%)</td>
</tr>
</tbody>
</table>

https://www.id-press.eu/mjms/index
Mean differences were also compared between groups using analysis of variant (ANOVA) (Table 2).

### Table 2: Means Difference of Leukocyte Subtypes

<table>
<thead>
<tr>
<th>Variables</th>
<th>Total Subjects</th>
<th>KPS 80-100 (n = 30)</th>
<th>KPS 50-70 (n = 34)</th>
<th>KPS 0-40 (n = 28)</th>
</tr>
</thead>
</table>
| Mean Leukocytes (x10⁶/µL) | 11.93 ± 9.96   | 11.48 ± 5.69         | 11.14 ± 5.38      | 13.05 ± 6.70      | 0.446
| Mean Neutrophil (%)       | 78.28 ± 9.74   | 73.21 ± 10.34        | 77.75 ± 8.86      | 80.74 ± 10.09     | 0.099
| Mean Neutrophil Absolute  | 9.57 ± 2.59    | 8.71 ± 5.01          | 8.83 ± 4.62       | 10.79 ± 6.06      | 0.303
| Mean Lymphocyte Percentage| 13.18 ± 7.68   | 17.03 ± 9.39         | 14.81 ± 7.89      | 9.84 ± 5.38       | 0.008
| Mean Lymphocyte Absolute  | 1.40 ± 0.74    | 1.66 ± 0.74          | 1.47 ± 0.67       | 1.23 ± 0.81       | 0.217
| Mean Monocyte Percentage  | 7.14 ± 3.21    | 7.04 ± 2.38          | 7.46 ± 3.42       | 6.81 ± 3.28       | 0.739
| Mean Monocyte Absolute     | 0.90 ± 0.63    | 0.92 ± 0.66          | 0.94 ± 0.67       | 0.84 ± 0.60       | 0.827
| Mean Basophil Percentage   | 0.31 ± 0.36    | 0.40 ± 0.38          | 0.36 ± 0.42       | 0.21 ± 0.28       | 0.202
| Mean Basophil Absolute     | 0.03 ± 0.03    | 0.04 ± 0.05          | 0.03 ± 0.03       | 0.03 ± 0.03       | 0.424
| Mean Neutrophil- Lymphocyte Ratio (NLR) | 8.23 ± 5.05 | 5.62 ± 2.94 | 6.67 ± 4.02 | 10.82 ± 5.72 | 0.001

*Significant p-value (<0.05).

Each variable was then correlated with KPS using Pearson's correlation test on the nature of normal distribution of the samples as seen in Table 3.

### Table 3: Pearson's Correlation of Leukocytes Subtypes and KPS

<table>
<thead>
<tr>
<th>Leukocytes Subtype</th>
<th>KPS 80-100 (n = 30)</th>
<th>KPS 50-70 (n = 34)</th>
<th>KPS 0-40 (n = 28)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Neutrophils (x10⁶/µL)</td>
<td>0.569</td>
<td>0.262</td>
<td>0.042</td>
</tr>
<tr>
<td>Neutrophil Percentage (%)</td>
<td>0.240</td>
<td>0.042</td>
<td>0.375</td>
</tr>
<tr>
<td>Neutrophil Absolute (x10⁶/µL)</td>
<td>0.010</td>
<td>0.375</td>
<td>0.383</td>
</tr>
<tr>
<td>Lymphocyte Percentage (%)</td>
<td>0.383</td>
<td>0.001</td>
<td>0.265</td>
</tr>
<tr>
<td>Lymphocyte Absolute (x10⁶/µL)</td>
<td>0.265</td>
<td>0.024</td>
<td>0.405</td>
</tr>
<tr>
<td>Monocyte Percentage (%)</td>
<td>0.100</td>
<td>0.405</td>
<td>0.121</td>
</tr>
<tr>
<td>Monocyte Absolute (x10⁶/µL)</td>
<td>0.121</td>
<td>0.311</td>
<td>0.188</td>
</tr>
<tr>
<td>Basophil Percentage (%)</td>
<td>0.188</td>
<td>0.115</td>
<td>0.153</td>
</tr>
<tr>
<td>Basophil Absolute (x10⁶/µL)</td>
<td>0.153</td>
<td>0.200</td>
<td>-0.432</td>
</tr>
<tr>
<td>Neutrophil-to- Lymphocyte Ratio (NLR)</td>
<td>-0.432</td>
<td>&lt; 0.001</td>
<td></td>
</tr>
</tbody>
</table>

*Significant p-value (<0.05).

### Discussion

On the mean differences and correlational analysis, we found significant differences on mean lymphocyte percentage between symptomatic group (KPS 80-100), symptomatic with assistance group (KPS 50-70) and symptomatic with bed confinement group (KPS 0-40), (7.03 ± 9.39 vs 14.81 ± 7.89% vs 9.84 ± 5.38%, respectively) with Pearson correlation product moment of lymphocyte percentage and KPS show rather weak but significant positive correlation (r = 0.383; p = 0.001) [24].

Although we were not getting any ANOVA's significant differences on mean neutrophile percentage and mean lymphocyte absolute, there are weak significant negative correlation on neutrophile percentage and KPS (r = -0.240, p = 0.042) also on lymphocyte absolute and KPS show weak significant positive correlation on lymphocyte absolute and KPS (r = 0.265, p = 0.024). This view was in line with my review on multiple tumour progression correlated with tumour-infiltrating neutrophils, as elevated blood neutrophils proven to be a poor prognostic factor for functional outcome [25] also stated that circulating tumour-infiltrating-lymphocyte number elevation also has correlated with favourable clinical outcomes in subsets of human cancer [26].

On NLR variable, we found significant mean differences percentage between symptomatic group (KPS 80-100), symptomatic with assistance group (KPS 50-70) and symptomatic with bed confinement group (KPS 0-40) (5.62 ± 2.94 vs 6.87 ± 4.02 vs 10.82 ± 5.72, respectively) and also significant moderate negative correlation between NLR and KPS (r = -0.432, p < 0.001). Although applied on different population (Glioblastoma Multiforme/GBM), NLR also has been studied by Alexiou et al., resulting on NLR > 4.7 was associated with decreased survival time (11 vs 18.7 months, p = 0.01) [7].

Templet et al., allege that high NLR has an adverse effect on overall survival (OS) in many solid tumours [27], and this result was also agreeable with the previous study by Serdarevic et al., [28] on BM and non-BM population of non-small-cell lung cancer (NSCLC) which conclude higher NLR mean on BM group and more progressive condition (6.05 vs 4.6; p = 0.023) [28]. This was also confirmed by another prognostic study of NLR that elevated preoperative NLR is a predictor of worse survival after BM resection (OS 14 month for NLR < 5 and 5 months for NLR ≥ 5; p = 0.001) [27, 29].

In conclusion, increased lymphocyte absolute and lymphocyte percentage correlated significantly with better KPS, while elevated neutrophils percentage and increased NLR show significant correlation with worse outcome of BM patients.

### References

5. Erez N, Coussens LM. Leukocytes as paracrine regulators of...


